A Cytotoxic 5a,8a-Epidioxysterol from a Soft Coral Sinularia Species

Jyh-Horng Sheu,* Kuie-Chi Chang, and Chang-Yih Duh

Department of Marine Resources, National Sun Yat-Sen University, Kaohsiung, Taiwan 804, Republic of China

Received August 9, 1999

A new sterol, (22R,23R,24R)- 5α , 8α -epidioxy-22,23-methylene-24-methylcholest-6-en- 3β -ol (1), as well as two known sterols, numersterol A (2) and pregnenolone (3), have been isolated from a soft coral *Sinularia* sp. The structure of metabolite 1 was determined by spectral analysis. Cytotoxicity of sterols 1-3 toward various cancer cell lines is also reported.

Soft corals have been well-recognized as marine organisms containing large quantities of secondary metabolites that exhibit varying degrees of biological activities.¹ Several cytotoxic compounds have been isolated previously from soft corals collected along the coast of Taiwan.²⁻⁸ In connection with our continuing investigations of bioactive substances from marine organisms, a soft coral Sinularia species was selected for study, as the EtOAc extract of this organism was found to exhibit significant cytotoxicity against P-388 (mouse lymphocytic leukemia) and KB (human nasopharyngeal carcinoma) cells. Initial study of the crude extract of this organism has led to the isolation of a novel cytotoxic hydroperoxysterol and five known compounds.9 Further investigation on the chemical constituents of this organism resulted in the isolation of a new cytotoxic sterol, (22R,23R,24R)-5a,8a-epidioxy-22,23-methylene-24-methylcholest-6-en- 3β -ol (1), and two known cytotoxic sterols, numersterol A (2) and pregnenolone (3).

Compound **1** was isolated from the fraction eluted with hexanes-EtOAc (3:1) as a white powdery solid, mp 159-160 °C, $[\alpha]^{26}_{D}$ +35° (c 0.1, CHCl₃). The HREIMS of 1 established a molecular formula of C₂₉H₄₆O₃, implying seven degrees of unsaturation. This sterol was further recognized as a 5α , 8α -epidioxy sterol by the presence of the characteristic signals for H-6 and H-7 at δ 6.25 (d, J =8.4 Hz) and 6.51 (d, J = 8.4 Hz), respectively, in the ¹H NMR spectrum.¹⁰ Four protons appeared at δ 0.13 (2H, m), 0.30 (1H, m), and 0.54 (1H, m) ppm, indicating the presence of a cyclopropyl group in the molecule. Two singlets, which appeared at δ 0.76 (3H) and 0.90 (3H) ppm, were attributed to C-18 and C-19 methyls, respectively. Four doublets at δ 0.86 (3H, J = 6.9 Hz), 0.88 (3H, J = 6.9 Hz), and 0.91 (6H, J = 6.3 Hz) ppm were due to the presence of C-28, C-27, C-21, and C-26 methyls, respectively. The above data suggested that 1 is a peroxysteroid containing a 22,23methylene-24-methyl moiety in the side chain. By comparison of the proton shifts of H₃-21, H₃-26, H₃-27, and H₃-28 with those of the four synthetic demethylgorgosterol isomers,¹¹ it was suggested that the stereochemistry of **1** at side chain should be assigned as 22R, 23R, and 24R(Figure 1). The assignment of the carbon shifts of 1 (Table 1) was based on the comparison of these data with those of the tetracyclic system of 5α , 8α -epidioxyergost-6-en- 3β ol¹² and those of the side chain carbons of stoloniferone*d*.¹³ The presence of a peroxide was further confirmed by the mass fragment ion, which showed peak at m/z 410 $[M - O_2]^+$, presumably *via* a retro-Diels-Alder fragmentation. Based on the above data, the structure of 1 was then

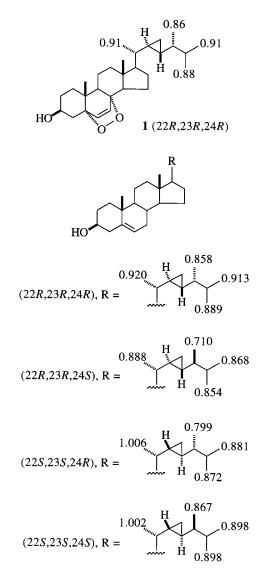


Figure 1. ¹H NMR Chemical shifts of the side-chain methyl groups of **1** and synthetic isomers of demethylgorgosterols.¹¹

established as (22R,23R,24R)-22,23-methylene-24-methylcholest-6-en-5 α ,8 α -epidioxy-3 β -ol. It was finally found that **1** is the deacetyl derivative of a known steroid, **4**.¹⁴ A structurally similar compound, **5**, isolated previously from a *Lobophytum* sp. soft coral, has been designated as 22,23methylene-24-methylcholest-6-en-5 α ,8 α -epidioxy-3 β -ol.¹⁵ However, the stereochemistry of **5** at C-22, C-23, and C-24 has not been characterized, and the ¹³C NMR data were not assigned and showed some discrepancies with those of

^{*} To whom correspondence should be addressed. Tel.: 886-7-5252000, ext. 5030. Fax: 886-7-5255020. E-mail: sheu@mail.nsysu.edu.tw.

Table 1. ¹³C NMR Chemical Shifts of Sterols 1-3

		compound	
position	1 ^a	2^{b}	3 ^c
C-1	39.3	75.9	38.3
C-2	30.1	39.2	32.5
C-3	66.5	64.8	71.1
C-4	51.1	41.9	42.3
C-5	79.5	79.2	142.4
C-6	130.8	76.4	121.4
C-7	135.4	35.3	32.8
C-8	82.2	31.5	32.6
C-9	34.7	41.5	52.6
C-10	36.9	44.0	37.4
C-11	20.8	21.7	21.9
C-12	39.6	41.3	39.8
C-13	44.8	41.7	44.4
C-14	51.3	57.6	57.7
C-15	28.4	25.3	23.5
C-16	23.4	29.3	39.6
C-17	57.8	57.6	64.0
C-18	12.4	12.6	13.6
C-19	18.5	17.3	19.9
C-20	39.5	34.9	208.5
C-21	19.1	19.1	31.5
C-22	24.2	36.0	
C-23	25.1	32.1	
C-24	45.0	157.9	
C-25	32.8	37.0	
C-26	18.1	22.4	
C-27	20.6	22.3	
C-28	15.7	106.9	
C-29	10.5		

^{*a*} The chemical shifts were determined at 75 MHz in CDCl₃. ^{*b*} 75 MHz in CD₃OD. ^{*c*} 75 MHz in acetone-*d*₆, respectively. The values are in ppm downfield from TMS.

Table 2. Cytotoxic Data of Sterols 1–3^{*a*}

	cell lines ED ₅₀ (µg/mL)			
compound	P-388	KB	A549	HT-29
1	0.4	2.1	2.7	1.4
2	8.3	1.9	10.8	1.5
3	7.8	>50	8.6	0.7

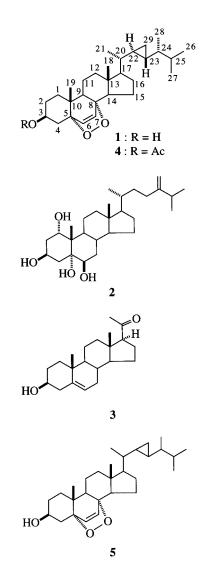
^{*a*} For significant activity of pure compounds, an ED₅₀ value of \leq 4.0 µg/mL is required. See Geran et al.²⁰

1 and the related compounds,^{12,13} particularly at C-29, the cyclopropyl methylene carbon, which should show a peak at about δ 10.5 ppm. In contrast, the most upfield signal of the eight methylene carbons of **5** was reported to appear at δ 23.3 ppm. Thus, peroxysterol **1** is regarded as a new compound.

The previously known compounds, numersterol A (**2**) and pregnenolone (**3**), were identified by their physical (mp and optical rotation) and spectral [IR, MS, ¹H and ¹³C NMR (Table 1)] data. The polyhydroxylated sterol **2** was isolated previously from the South China Sea soft coral *Sinularia numerosa*.¹⁶ The occurrence of pregnenolone (**3**) in a sponge *Haliclona rubens* has been reported;¹⁷ however, this is the first observation for the existence of **3** in a soft coral.

To find the future biomedical potential for the above steroids, cytotoxicity testing was performed. The cytotoxicity of these compounds toward a limited panel of cancer cell lines is shown in Table 2. A sterol with a structure similar to 1, $5\alpha,8\alpha$ -epidioxyergost-6-en-3 β -ol, has been shown to exhibit cytotoxicity toward MCF-7 breast and Walker 256 carcinomasarcoma cell lines.¹² Our present study also shows that 1 exhibited significant cytotoxicity toward the growth of P-388, KB, A549, and HT-29 cells. The results of these two investigations indicated that steroids containing a $5\alpha,8\alpha$ -epidioxy functional group may warrant further antitumor studies in the future. Compound





2 exhibited selective cytotoxicity against KB and HT-29 cells. Compound **3** also was found to show significant activity exhibiting the growth of HT-29 cells.

Experimental Section

General Experimental Procedures. Melting points were determined using a Fisher–Johns melting points apparatus and were uncorrected. Optical rotations were measured on a JASCO DIP-1000 digital polarimeter. The IR spectra were measured on a Horiba FT-720 infrared spectrophotometer. ¹H and ¹³C NMR spectra were recorded on a Varian VXR-300/5 FT NMR spectrometer at 300 and 75 MHz, respectively, in CDCl₃ using TMS as internal standard, unless otherwise indicated. EIMS spectra were obtained with a VG QUATTRO GC–MS spectrometer. HREIMS spectra was recorded on a JMX-HX 110 mass spectrometer. Si gel 60 (Merck, 230–400 mesh) was used for column chromatography. Precoated Si gel plates (Merck, Kieselgel 60 F₂₅₄, 0.20 mm) were used for analytical TLC.

Animal Material. The soft coral *Sinularia* sp. was collected by hand using scuba at the South Bay, Kenting, located in the southernmost tip of Taiwan, in June 1995, at a depth of 4 m and was immediately stored in a freezer until extraction. The colony morphology and the shape of the sclerites in both the surface and interior of the lobe indicate that the sample is a soft coral of the genus *Sinularia*.¹⁸ A voucher specimen was deposited in the Department of Marine Resources, National Sun Yat-Sen University (specimen no. KTSC-105).

Extraction and Separation. The soft coral (3.3 kg fresh wt) was collected and freeze-dried. The freeze-dried material

(0.8 kg) was minced and extracted exhaustively with EtOAc (8 L \times 5). The organic extract was evaporated to dryness, and the oily residue (8.3 g) was found to exhibit cytotoxcicity against the P-388 cell line, with an ED₅₀ of 12.8 μ g/mL, and the KB cell line, with an ED₅₀ of 15.4 μ g/mL. The extract was separated by Si gel column chromatography. Sterol 3 was eluted with hexanes-EtOAc (4:1), 1 with hexanes-EtOAc (3: 1), and 2 with EtOAc-EtOH (3:1).

(22R,23R,24R)-5a,8a-Epidioxy-22,23-methylene-24methylcholest-6-en- 3β -ol (1): white powder (1.7 mg); mp 159–160 °C; $[\alpha]^{26}_{D}$ +35° (*c* 0.1, CHCl₃); IR (KBr) ν_{max} 3380, 1646, 1062 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 0.13 (2H, m, H2-29), 0.30 (1H, m, H-22a), 0.54 (1H, m, H-23\beta), 0.54 (1H, m, H-24), 0.76 (3H, s, H₃-18), 0.86 (3H, d, J = 6.9 Hz, H₃-28), 0.88 (3H, d, J = 6.9 Hz, H₃-27), 0.90 (3H, s, H₃-19), 0.91 (6H, d, J = 6.3 Hz, H₃-21 and H₃-26), 3.98 (1H, m, H-3), 6.25 (1H, d, J = 8.4 Hz, H-6), 6.51 (1H, d, J = 8.4 Hz, H-7); ¹³C NMR, see Table 1; EIMS (30 eV) m/z (rel int) 442 (1, M⁺), 424 (3), 410 (7), 303 (1), 301 (3), 271 (3), 269 (1), 253 (5), 251 (9), 69 (100); HREIMS m/z 442.3472 (calcd for C₂₉H₄₆O₃ 442.3435).

Numersterol A (2): white powder (18.5 mg); mp 296-298 °C; $[\alpha]^{26}_{D}$ +5° (*c* 0.3, MeOH); IR (KBr) ν_{max} 3330, 1678, 1070, 890 cm $^{-1};$ 1H NMR (300 MHz, CD₃OD) δ 0.69 (3H, s, H₃-18), 0.95 (3H, d, J = 6.3 Hz, H₃-21), 1.02 (6H, d, J = 6.9 Hz, H₃-26 and H₃-27), 1.11 (3H, s, H₃-19), 3.49 (1H, br s, H-6a), 3.61 (1H, br s, OH), 3.89 (1H, t, J = 3.9 Hz, H-1 β), 4.30–4.50 (1H, m, H-3 α), 4.65 (1H, s, H-28), 4.71 (1H, s, H-28); ¹³C NMR, see Table 1; EIMS (30 eV) m/z (rel int) 433 [3, (M - CH₃)⁺], 412 (8), 397 (5), 346 (12), 321 (18), 69 (100).

Pregnenolone (3): white powder (10.2 mg); mp 192–193 °C; $[\alpha]^{26}_{D} + 27^{\circ}$ (c 0.3, CHCl₃); IR (KBr) ν_{max} 3410, 1710, 1650 cm⁻¹; ¹H NMR (300 MHz, acetone- d_6) δ 0.62 (3H, s, H₃-18), 1.01 (3H, s, H₃-19), 2.06 (3H, s, H₃-21), 2.61 (1H, t, J = 9.0Hz, H-17), 3.42 (1H, m, H-3), 3.80 (1H, d, J = 4.8 Hz, OH), 5.33 (1H, d, J = 4.5 Hz, H-6); ¹³C NMR, see Table 1; EIMS (30 eV) m/z (rel int) 316 (2, M⁺), 298 (2), 283 (2), 43 (100).

Cytotoxicity Testing. KB and P-388 cells were kindly provided by Prof. J. M. Pezzuto, University of Illinois at Chicago; A549 (human lung adenocarcinoma) and HT-29 (human colon adenocarcinoma) were purchased from the American Type Culture Collection. The cytotoxic activities of tested compounds 1-3 were assayed by a modification of the MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide] colorimetric method.¹⁹ The cultured cells were treated at eight concentrations of pure test compounds ranging from 0.00064 to 50 μ g/mL. All assays were performed in triplicate. The results were expressed as a percentage, relative to control incubations, and the effective dose required to inhibit cell growth by 50% (ED₅₀) was determined.

Acknowledgment. This work was supported by grants from the National Science Council of the Republic of China (NSC-87-2113-M-110-015 and NSC-88-2113-M-110-011) awarded to J.-H. Sheu. We thank Prof. Keryea Soong (Institute of Marine Biology, National Sun Yat-Sen University) for identification of the specimen.

References and Notes

- (1) Faulkner, D. J. Nat. Prod. Rep. 1999, 16, 155-198, and previous reports in this series
- (2) Hou, R.-S.; Duh, C.-Y.; Chiang, M. Y.; Lin, C.-N. J. Nat. Prod. 1995, 58, 1126-1130.
- (3) Duh, C.-Y.; Hou, R.-S. J. Nat. Prod. 1996, 59, 595-598.
 (4) Sheu, J.-H.; Yeh, T.-H. J. Chin. Chem. Soc. 1991, 38, 397-399.
 (5) Duh, C.-Y.; Wang, S.-K.; Tseng, H.-K.; Sheu, J.-H.; Chiang, M. Y. J. Nat. Prod. 1998, 61, 844-847.
- Sheu, J.-H.; Sung, P.-J.; Huang, L.-H.; Lee, S.-F.; Wu, T.; Chang, B.-(6)Y.; Duh, C.-Y.; Fang, L.-S.; Soong, K.; Lee, T.-J. J. Nat. Prod. 1996, 59. 935-938.
- 59, 935-938.
 (7) Sheu, J.-H.; Sung, P.-J.; Cheng, M.-C.; Liu, H.-Y.; Fang, L.-S.; Duh, C.-Y.; Chiang, M. Y. J. Nat. Prod. 1998, 61, 602-608.
 (8) Sung, P.-J.; Su, J.-H.; Wang, G.-H.; Lin, S.-F.; Duh, C.-Y.; Sheu, J.-H. J. Nat. Prod. 1999, 62, 457-463.
- (9) Sheu, J.-H.; Chang, K.-C.; Sung, P.-J.; Duh, C.-Y.; Shen, Y.-C. J. Chin.
- (a) Shet, 3.-11, Chang, N.-C., Sung, I.-O., Dun, C.-11, Shen, T.-C. J. Chin. Chem. Soc. 1999, 46, 253–257.
 (10) Gunatilaka, A. A. L.; Gopichand, Y.; Schmitz, F. J.; Djerassi, C. J. Org. Chem. 1981, 46, 3860–3866.
 (11) Blanc, P.-A.; Djerassi, C. J. Am. Chem. Soc. 1980, 102, 7113–7114.
 (12) Kahlos, K.; Kangas, L.; Hiltunen, R. Planta Med. 1989, 55, 389–200.
- 390. (13) Kobayashi, M.; Lee, N. K.; Son, B. W.; Yanagi, K.; Kyogoku, Y.;
- Kitagawa, I. Tetrahedron Lett. 1984, 25, 5925-5928. Anjanevulu, A. S. R.; Sagar, K. S.; Venugopal, M. J. R. V. *Tetrahedron* **1995**, *51*, 10997–11010. (14)
- Subrahmanyam, C.; Rao, C. V.; Kulatheeswaran, R. Indian J. Chem. (15)**1995**, *34B*, 1114–1115.
- (16)
- Su, J.; Yu, X.; Zeng, L. J. Nat. Prod. **1989**, *52*, 934–940. Ballantine, J. A.; Williams, K. Tetrahedron Lett. **1977**, 1547–1550. (17)
- Bayer, F. M. Proc. Biol. Soc. Wash. 1981, 94, 902-947. (18)
- (19) Mosmann, T. J. Immunol. Methods 1983, 65, 55-63.
- (20)Geran, R. I.; Greenberg, N. H.; MacDonald, M. M.; Schumacher, A. M.; Abbott, B. J. Cancer Chemother. Rep. 1972, (3), 1-91.

NP9903954